16 week protozoa

Note: The culturing conditions below are not necessarily the optimal growth conditions for each strain, as much variation is found between strains, and cultures are not always kept in optimal growth conditions at CCAP for practical reasons. There may be more info in the individual strain data on the website.

On receipt of culture: If the culture vessel is very full on receipt and subculturing is not necessary immediately, we advise transferring half of the culture to a sterile container to provide air space.

ACDP Hazard Gp: 1 - Non pathogenic / non hazardous. Unlikely to cause human disease.

Culture Medium: Check individual strain data, strains in this group are grown in MW, ASWP, SES, or a combination. All are supplemented with a sterilised wheat grain to encourage growth of the bacteria the nanoflagellates feed on.
Media recipes can be found on our website: www.ccap.ac.uk/index.php/media-recipes/

Lighting: subdued light

Light Cycle: natural day/night cycle

Temperature: 15 degrees C; the following strains are kept at 20 degrees C:

CCAP 1576/1 *Phalansterium filosum*
CCAP 1921/1 *Filoreta turcica*
CCAP 1921/2 *Filoreta marina*
CCAP 1900/3 *Cafeteria roenbergensis*
CCAP 1953/1 *Nutomonas kenti*

Sub Interval: 16 weeks (may vary depending on environment)

Culture Vessel: petri dishes

Culture Method:

All strains are cultured within petri dishes that can be examined directly using an inverted microscope x40. Examine each plate before subculturing.

One new subculture is created from each petri dish unless density is low in which case two or more dishes can be subcultured.

Under a fume hood, label sterile petri dishes and fill up to half way with sterilised liquid medium.

Use a sterile pipette to transfer 1-2ml of fresh medium into the older culture. Squeeze the bulb of the pipette while moving it across the base of the dish to resuspend the organisms, then suck up a couple of ml in a zig-zag pattern and transfer to dish of fresh medium. Sterilise forceps and add wheat grain (pre-sterilise wheat by boiling in distilled water). Seal dish with parafilm.

Use strict aseptic techniques throughout and if possible carry out all subculturing within a laminar flow cabinet.